

## Biotransformation of Rutin to Quercetin by Human Gut Bacteria and Its Effect on Rutin Bioavailability

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**Abstract** -Rutin is a flavonol glycoside containing quercetin and the rutinose. Rutin exert a wide range of biological benefits for human being. However, it is poorly absorbed by small intestine and most reaches the colon. Human gut bacteria play important role in biotransformation of rutin to quercetin. The biological activities of flavonoids and their metabolic role after consumption depends on their chemical structure and absorption. For instance, quercetin is the aglycone form of rutin with higher bioavailability, absorption in small intestine than rutin. The microbial biotransformation not only improves biological activities of plant glycosides but also improves their bioavailability. Gut microbiota was shown to improve rutin colonic absorption after its hydrolysis by  $\beta$ -glucosidase and  $\alpha$ -l-rhamnosidase. The aim of the present review is to give updates on biological properties of rutin, biotransformation of rutin to quercetin by gut bacteria strains and its effect on rutin bioavailability.

**Keywords**:- Bioavailability, microbial biotransformation, rutin, quercetin

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### I. Introduction

Flavonoids are a group of bioactive compounds that are extensively found in foodstuffs of plant origin. The flavonoids are  $\alpha$  or  $\beta$  glycoside such as flavonoid glucosides, galactosides, rhamnosides, arabinosides and rutinoides [1-5]. One of these flavonoids, rutin (quercetin-3-O-rutinoside) is a flavonol glycoside composed of quercetin and rutinose [6, 7]. The plant sources of rutin includes vegetables and fruits such as tea, green asparagus, ruta graveolens, onions, buckwheat, wine, eucalyptus spp., apples, and sophora japonica as well as berries [8-12]. Rutin possess antioxidant properties, vasoprotective, anti-cancer, anti-hypercholesterolemia, anti-diabetic, anti-hypertensive, exerts renal protective effects [13-16] as well as reducing the risk of atherosclerosis [16]. Quercetin is the aglycone form of flavonoid glycosides such as rutin and quercitrin [17]. Quercetin was reported to have so many biological properties [18] including anti-osteoporosis, anti-inflammatory [19], anti-carcinogenic, anti-pulmonary and cardiovascular diseases, anti-aging [20], antiviral, antioxidant, and psychostimulant activities as well as immunological improvement [21].

However, the bioavailability of rutin is relatively low due to its low solubility which limits its practical applications [22-25]. In addition, glycosylation by rutinose limits the absorption of flavonoids in the small intestine [26]. Moreover, quercetin aglycone has a very high bioavailability [27], but rutin is absorbed comparatively slowly and most reaches the colon [28]. Therefore, biotransformation of polyphenols changes the structure of polyphenols to improve bioavailability and maintains their original bioactivity [29]. In order to exert biological effects, gut microbiota contributes a lot in the bioavailability and health effects of poorly absorbed polyphenols [30-32]. Glycosides reach microflora in colon where they are hydrolyzed by  $\beta$ -glucosidase from gut bacteria (Griffiths and Barrow 1972, Di Gioia, Bregola et al. 2010). Few studies have been focused on biotransformation of rutin to quercetin by gut bacteria. The aim of the present review is to give updates on biological properties of rutin, biotransformation of rutin to quercetin by gut bacteria strains and its effect on rutin bioavailability.

### II. Biological Properties Of Quercetin And Rutin

The biological effects are associated to the metabolites which succeed to reach the internal organs rather than their native compounds found in foods **Table 1** [33]. Moreover, microbial degradation of flavonoids in the human intestine beneficially affect human health.

For example, the antioxidant and antiproliferative potential of rutin deglycosylated with hesperidinase and naringinase were higher than that of rutin [34, 35]. Moreover, antioxidant activity of flavonoid measured as DPPH radical scavenging activity was found to be 62.6% for rutin and 89.8% for quercetin [35]. Quercetin exerts hepatoprotective [36], anti-inflammatory activities [37], strongest antioxidant activities compared to other flavonoids [38, 39], anti-skin damages [40], neuroprotective effects on brain injury [41], anti-apoptotic [42, 43], anti-aging activities [44], anti-colorectal lung metastasis [45], anti-breast cancer, prostate cancer activities [46].

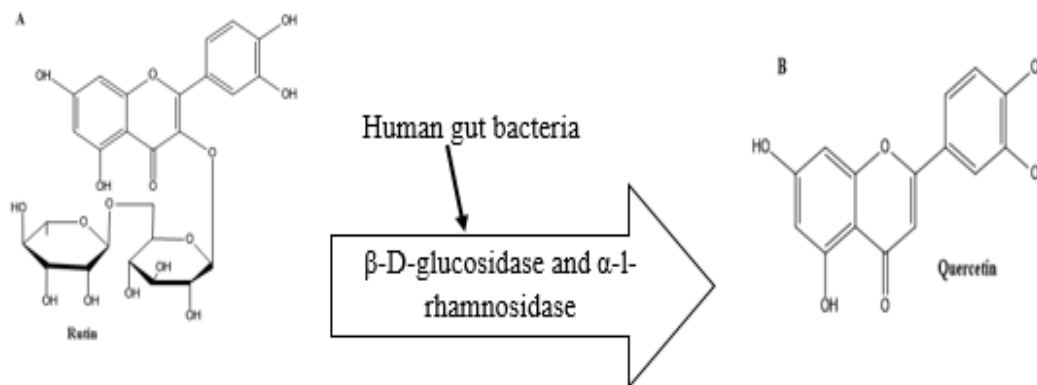
**Table 1. Biological properties of rutin**

Biological properties	References
Anti-Cancer	[35, 47-49]
Antioxidant	[50, 51]
Antiallergic	[52]
Anti-inflammatory	[53-55]
Hypolipidaemic	[56, 57]
Neuroprotective	[58-61]
Antihyperglycaemic	[62, 63]
Gastro-protective activity	[64]

### III. Biotransformation Of Rutin To Quercetin By Human Gut Bacteria

The microbial biotransformation of rutin to quercetin have been reported [65]. This results from hydrolysis of rutin by bacterial  $\alpha$ -rhamnosidases and  $\beta$ -glucosidases in the colon as shown in **Fig.1** [11, 66].

Rutin require hydrolysis by intestinal or microbial enzymes for their absorption in vivo [67, 68]. Some colonic bacteria including *Lactobacillus* spp., *Bacteroides* spp. and *Bifidobacterium* spp. can cleave glycosides by their  $\beta$ -glucosidases [69].



**Figure 1. Human gut bacteria enzymes converting rutin to quercetin**

Rutin is absorbed from the colon after deglycosylation [70]. The colonic microbiota break down rutin to release quercetin [71, 72]. During fermentation, kaempferol and quercetin glycosides were found to be transformed in their corresponding aglycones [73, 74]. Biotransformation can produce new compounds with potential biological activity [75, 76]. In addition, metabolism of phenolic compounds by microbiota can only occur after deglycosylation or deconjugation [77, 78].

Researches has shown that metabolites from dietary polyphenols might have greater biological activities than their parent compounds [28, 79]. For example the deglycosylation of flavonoid glycosides to their aglycones have been reported to improve also their antioxidant activity [3]. Microbial biotransformation, metabolic engineering and enzyme engineering have been reported for production of bioactive flavonoids [80, 81]. Furthermore, microbial fermentation is mostly used for biotransformation [82] due to its advantages as shown in **Table 2** [83, 84].

**Table 2. General advantages of microbial biotransformation**

Advantages	References
Improve the production yield	[83-85]
Microbial cells or enzymes can be immobilized and reused for many cycles	[29]
High specificity	[86-88]
Economic and environmental friendly	[89-91]

Plant metabolite deglycosylation was shown to improve the flavor and taste of fermented products [92]. Glycosidases from some microorganisms shown in **Table 3** ( $\alpha$ -l-rhamnosidase and  $\beta$ -glucosidase) hydrolyze the bonds between quercetin and rutinose, and between glucose and rhamnose[93].

**Table 3. Some microorganisms involved in biotransformation of rutin to quercetin**

Microorganism	Product	References
<i>Enterococcus avium</i> EFEL009	Quercetin	[94]
<i>Bacteroides uniformis</i> ATCC 8492T	Quercetin	[95]
<i>Lactobacillus plantarum</i> , <i>Lactobacillus acidophilus</i>	Quercetin -3-O-glucoside	[96]
<i>Bacteroides sp. 22 and Bacteroides sp. 45</i>	Quercetin -3-O-glucoside	[97]

*Fusobacterium K-60* and *Bacteroides JY-6* by could degrade rutin to lose sugars by  $\beta$ -glucosidase and  $\alpha$ -rhamnosidase. Lactobacilli are probiotics and among the predominant members of the intestinal bacteria[98]. The presence of *Lactobacillus plantarum* in the gut microflora was reported to enhance flavonoid bioavailability [99]. Lactobacilli convert rutin and hesperidin into their aglycone forms and contribute to their bioavailability and absorption [100-102].*Lactobacillus buchneri* and *Lactobacillus acidophilus* were able to convert wheat flavonoid glycosides into aglyconic forms to increase their absorption [101]. *Lactobacillus* is estimated to constitute 6% of the total bacterial cell numbers in the human duodenum and approximately 0.3% of all bacteria in the colon[103]. The bacterial concentration in the small intestine is low. So *Lactobacillus* spp. should not be major contributor of rutin transformer in vivo. Due to long history of application as probiotic and in fermented food, it has the industrial potential to transform rutin in food production.

#### IV. Effect Of Biotransformation Rutin To Quercetinon Rutin Bioavailability

The bioavailability for intestinal absorption, interaction with target tissues and metabolism of bioactive polyphenols influence their health promoting effects[104]. Flavonoids need to be consumed in a form that can be absorbed in the small intestine before their potential degradation [99]. More complex flavonoid glycosides are not absorbed by the small intestine and reach the colon [105].

For example, rutin is poorly absorbed by small intestine but mostly degraded by the gut bacteria after consumption [11, 106]. The enzymatic de-glycosylation of flavonoids has been reported to increase their bioavailability[107]. For instance, rutin deglycosylation by  $\beta$ -glucosidase from gut microbiota (Kim, Jung et al. 1998) was shown to improve its colonic absorption [108]. In addition, removal of rhamnose group by rhamnosidases improves also the bioavailability of flavonoids. The microbial biotransformation not only improves bioavailability but also provides more selectivity and less toxicity compared to chemical synthesis [31, 109-111].

Many aglycones and some of the monoglycosides are absorbed in the ileum [112]. Moreover, the study showed that bioavailability of quercetin from rutin is higher than that from quercetin aglycone in cows after oral application [113]. It has been reported that deglycosylation of quercetin glycosides by  $\beta$ -glucosidase, improves its absorption[114]. Therefore quercetin aglycone is easy for the small intestine uptake[115-117].

#### V. Conclusion

Rutin exerts so many biological effects including antioxidant, anti-cancer, anti-hypercholesterolemia, anti-diabetic, anti-hypertensive as well as renal protective effects. Unfortunately rutin absorption in small intestine is lower than that of quercetin poorly and a large amount reach the colon where it is degraded by human intestinal bacteria. Human gut bacteria hydrolyze rutin to quercetin by  $\alpha$ -l-rhamnosidase and  $\beta$ -glucosidase. In summary, health-promoting effects of rutin, biotransformation of rutin to quercetin and rutin bioavailability are greatly influenced by human gut bacteria strains. In addition, there very few studies done on probiotic bacteria strains involved in the biotransformation of rutin to quercetin. Further studies are needed to isolate and identify more human gut bacteria and probiotic strains which contribute to biotransformation of rutin to quercetin.

#### References

- [1]. R. Taheri, B.A. Connolly, M.H. Brand, B.W. Bolling, Underutilized Chokeberry (*Aronia melanocarpa*, *Aronia arbutifolia*, *Aronia prunifolia*) Accessions Are Rich Sources of Anthocyanins, Flavonoids, Hydroxycinnamic Acids, and Proanthocyanidins, *J. Agric. Food Chem.* 61(36) (2013) 8581-8588.
- [2]. R. Fang, N.C. Veitch, G.C. Kite, E.A. Porter, M.S.J. Simmonds, Enhanced Profiling of Flavonol Glycosides in the Fruits of Sea Buckthorn (*Hippophae rhamnoides*), *J. Agric. Food Chem.* 61(16) (2013) 3868-3875.
- [3]. J. Xiao, T.S. Muzashvili, M.I. Georgiev, Advances in the biotechnological glycosylation of valuable flavonoids, *Biotechnology Advances* 32(6) (2014) 1145-1156.

- [4]. M. Plaza, T. Pozzo, J. Liu, K.Z. Gulshan Ara, C. Turner, E. Nordberg Karlsson, Substituent Effects on in Vitro Antioxidizing Properties, Stability, and Solubility in Flavonoids, *J. Agric. Food Chem.* 62(15) (2014) 3321-3333.
- [5]. D. Del Rio, A. Rodriguez-Mateos, J.P.E. Spencer, M. Tognolini, G. Borges, A. Crozier, Dietary (Poly)phenolics in Human Health: Structures, Bioavailability, and Evidence of Protective Effects Against Chronic Diseases, *Antioxidants & Redox Signaling* 18(14) (2012) 1818-1892.
- [6]. S. Tranchimand, P. Brouant, G. Iacazio, The rutin catabolic pathway with special emphasis on quercetinase, *Biodegradation* 21(6) (2010) 833-859.
- [7]. R. Guo, P. Wei, W. Liu, Combined antioxidant effects of rutin and Vitamin C in Triton X-100 micelles, *Journal of Pharmaceutical and Biomedical Analysis* 43(4) (2007) 1580-1586.
- [8]. B. Gullón, T.A. Lú-Chau, M.T. Moreira, J.M. Lema, G. Eibes, Rutin: A review on extraction, identification and purification methods, biological activities and approaches to enhance its bioavailability, *Trends in Food Science & Technology* 67 (2017) 220-235.
- [9]. L.S. Chua, A review on plant-based rutin extraction methods and its pharmacological activities, *Journal of Ethnopharmacology* 150(3) (2013) 805-817.
- [10]. K.H. Kim, K.W. Lee, D.Y. Kim, H.H. Park, I.B. Kwon, H.J. Lee, Optimal recovery of high-purity rutin crystals from the whole plant of *Fagopyrum esculentum* Moench (buckwheat) by extraction, fractionation, and recrystallization, *Bioresource Technology* 96(15) (2005) 1709-1712.
- [11]. C. Manach, C. Morand, C. Demigné, O. Texier, F. Régéat, C. Rémésy, Bioavailability of rutin and quercetin in rats, *FEBS Letters* 409(1) (1997) 12-16.
- [12]. J.M. Drinkwater, R. Tsao, R. Liu, C. Defelice, D.J. Wolyn, Effects of cooking on rutin and glutathione concentrations and antioxidant activity of green asparagus (*Asparagus officinalis*) spears, *Journal of Functional Foods* 12 (2015) 342-353.
- [13]. C. Carrasco-Pozo, M.L. Mizgier, H. Speisky, M. Gotteland, Differential protective effects of quercetin, resveratrol, rutin and epigallocatechin gallate against mitochondrial dysfunction induced by indomethacin in Caco-2 cells, *Chemico-Biological Interactions* 195(3) (2012) 199-205.
- [14]. S.K. Panchal, H. Poudyal, T.V. Arumugam, L. Brown, Rutin Attenuates Metabolic Changes, Nonalcoholic Steatohepatitis, and Cardiovascular Remodeling in High-Carbohydrate, High-Fat Diet-Fed Rats, *The Journal of Nutrition* 141(6) (2011) 1062-1069.
- [15]. S. Sharma, A. Ali, J. Ali, J.K. Sahni, S. Baboota, Rutin: therapeutic potential and recent advances in drug delivery, *Expert Opinion on Investigational Drugs* 22(8) (2013) 1063-1079.
- [16]. A. Korkmaz, D. Kolankaya, Protective Effect of Rutin on the Ischemia/Reperfusion Induced Damage in Rat Kidney, *Journal of Surgical Research* 164(2) (2010) 309-315.
- [17]. I. Erlund, T. Kosonen, G. Alftan, J. Mäenpää, K. Perttunen, J. Kenraali, J. Parantainen, A. Aro, Pharmacokinetics of quercetin from quercetin aglycone and rutin in healthy volunteers, *European Journal of Clinical Pharmacology* 56(8) (2000) 545-553.
- [18]. M. Russo, C. Spagnuolo, I. Tedesco, S. Bilotto, G.L. Russo, The flavonoid quercetin in disease prevention and therapy: Facts and fancies, *Biochemical Pharmacology* 83(1) (2012) 6-15.
- [19]. J.H. Jung, J.I. Kang, H.S. Kim, Effect of quercetin on impaired immune function in mice exposed to irradiation, *Nutrition research and practice* 6(4) (2012) 301-7.
- [20]. A.W. Boots, G.R.M.M. Haenen, A. Bast, Health effects of quercetin: From antioxidant to nutraceutical, *European Journal of Pharmacology* 585(2-3) (2008) 325-337.
- [21]. Y. Li, J. Yao, C. Han, J. Yang, M. Chaudhry, S. Wang, H. Liu, Y. Yin, Quercetin, Inflammation and Immunity, *Nutrients* 8(3) (2016) 167.
- [22]. K. Miyake, H. Arima, F. Hirayama, M. Yamamoto, T. Horikawa, H. Sumiyoshi, S. Noda, K. Uekama, Improvement of solubility and oral bioavailability of rutin by complexation with 2-hydroxypropyl-beta-cyclodextrin, *Pharm. Dev. Technol.* 5(3) (2000) 399-407.
- [23]. R. Kamel, D.M. Mostafa, Rutin nanostructured lipid cosmeceutical preparation with sun protective potential, *Journal of Photochemistry and Photobiology B: Biology* 153 (2015) 59-66.
- [24]. M.L. Calabrò, S. Tommasini, P. Donato, R. Stancanelli, D. Raneri, S. Catania, C. Costa, V. Villari, P. Ficarra, R. Ficarra, The rutin/ $\beta$ -cyclodextrin interactions in fully aqueous solution: spectroscopic studies and biological assays, *Journal of Pharmaceutical and Biomedical Analysis* 36(5) (2005) 1019-1027.
- [25]. R. Mauludin, R.H. Müller, C.M. Keck, Development of an oral rutin nanocrystal formulation, *International Journal of Pharmaceutics* 370(1-2) (2009) 202-209.
- [26]. M. Mueller, B. Zartl, A. Schleritzko, M. Stenzl, H. Viernstein, F.M. Unger, Rhamnosidase activity of selected probiotics and their ability to hydrolyse flavonoid rhamnoglucosides, *Bioprocess and Biosystems Engineering* 41(2) (2018) 221-228.
- [27]. V. Crespy, C. Morand, C. Besson, C. Manach, C. Demigne, C. Remesy, Quercetin, but not Its Glycosides, Is Absorbed from the Rat Stomach, *J. Agric. Food Chem.* 50(3) (2002) 618-621.
- [28]. M. Monagas, M. Urpi-Sarda, F. Sanchez-Patan, R. Llorach, I. Garrido, C. Gomez-Cordoves, C. Andres-Lacueva, B. Bartolome, Insights into the metabolism and microbial biotransformation of dietary flavan-3-ols and the bioactivity of their metabolites, *Food & Function* 1(3) (2010) 233-253.
- [29]. A. Gupta, L.D. Kagliwal, R.S. Singhal, Chapter Four - Biotransformation of Polyphenols for Improved Bioavailability and Processing Stability, in: H. Jeyakumar (Ed.), *Advances in Food and Nutrition Research*, Academic Press 2013, pp. 183-217.
- [30]. M.P. Gonthier, C. Remesy, A. Scalbert, V. Cheynier, J.M. Souquet, K. Poutanen, A.M. Aura, Microbial metabolism of caffeic acid and its esters chlorogenic and caftaric acids by human faecal microbiota in vitro, *Biomedicine & Pharmacotherapy* 60(9) (2006) 536-540.
- [31]. A. Chandrasekara, F. Shahidi, Bioaccessibility and antioxidant potential of millet grain phenolics as affected by simulated in vitro digestion and microbial fermentation, *Journal of Functional Foods* 4(1) (2012) 226-237.
- [32]. J.C. Espin, A. Gonzalez-Sarrias, F.A. Tomas-Barberan, The gut microbiota: A key factor in the therapeutic effects of (poly) phenols, *Biochemical Pharmacology* 139 (2017) 82-93.
- [33]. D. Del Rio, L.G. Costa, M.E.J. Lean, A. Crozier, Polyphenols and health: What compounds are involved?, *Nutrition, Metabolism and Cardiovascular Diseases* 20(1) (2010) 1-6.
- [34]. K.C.F. Araújo, E.M. de M.B. Costa, F. Pazini, M.C. Valadares, V. de Oliveira, Bioconversion of quercetin and rutin and the cytotoxicity activities of the transformed products, *Food and Chemical Toxicology* 51 (2013) 93-96.
- [35]. M.E.M.B. de Araújo, Y.E. Moreira Franco, T.G. Alberto, M.A. Sobreiro, M.A. Conrado, D.G. Priolli, A.C.H. Frankland Sawaya, A.L.T.G. Ruiz, J.E. de Carvalho, P. de Oliveira Carvalho, Enzymatic de-glycosylation of rutin improves its antioxidant and antiproliferative activities, *Food Chemistry* 141(1) (2013) 266-273.
- [36]. S. Miltonprabu, M. Tomczyk, K. Skalicka-Woźniak, L. Rastrelli, M. Daglia, S.F. Nabavi, S.M. Alavian, S.M. Nabavi, Hepatoprotective effect of quercetin: From chemistry to medicine, *Food and Chemical Toxicology* 108 (2017) 365-374.

- [37]. C.-F. Lin, Y.-L. Leu, S.A. Al-Suwayeh, M.-C. Ku, T.-L. Hwang, J.-Y. Fang, Anti-inflammatory activity and percutaneous absorption of quercetin and its polymethoxylated compound and glycosides: The relationships to chemical structures, *European Journal of Pharmaceutical Sciences* 47(5) (2012) 857-864.
- [38]. T. Hatahet, M. Morille, A. Hommoss, C. Dorandeu, R.H. Müller, S. Bégu, Dermal quercetin smartCrystals®: Formulation development, antioxidant activity and cellular safety, *European Journal of Pharmaceutics and Biopharmaceutics* 102 (2016) 51-63.
- [39]. D.A. Valério, S.R. Georgetti, D.A. Magro, R. Casagrande, T.M. Cunha, F.T.M.C. Vicentini, S.M. Vieira, M.J.V. Fonseca, S.H. Ferreira, F.Q. Cunha, W.A. Verri, Quercetin Reduces Inflammatory Pain: Inhibition of Oxidative Stress and Cytokine Production, *J. Nat. Prod.* 72(11) (2009) 1975-1979.
- [40]. D. Liu, H. Hu, Z. Lin, D. Chen, Y. Zhu, S. Hou, X. Shi, Quercetin deformable liposome: Preparation and efficacy against ultraviolet B induced skin damages in vitro and in vivo, *Journal of Photochemistry and Photobiology B: Biology* 127 (2013) 8-17.
- [41]. B. Chen, J. Park, J. Ahn, J. Cho, I. Kim, J. Lee, M.-H. Won, C.-H. Lee, I. Hwang, J.-D. Kim, I. Kang, J. Cho, B. Shin, Y. Kim, Y. Lee, S. Park, Pretreated quercetin protects gerbil hippocampal CA1 pyramidal neurons from transient cerebral ischemic injury by increasing the expression of antioxidant enzymes, *Neural Regeneration Research* 12(2) (2017) 220-227.
- [42]. T. Yang, B. Kong, J.-W. Gu, Y.-Q. Kuang, L. Cheng, W.-T. Yang, X. Xia, H.-F. Shu, Anti-apoptotic and Anti-oxidative Roles of Quercetin After Traumatic Brain Injury, *Cellular and Molecular Neurobiology* 34(6) (2014) 797-804.
- [43]. R.-Q. Yao, D.-S. Qi, H.-L. Yu, J. Liu, L.-H. Yang, X.-X. Wu, Quercetin Attenuates Cell Apoptosis in Focal Cerebral Ischemia Rat Brain Via Activation of BDNF-TrkB-PI3K/Akt Signaling Pathway, *Neurochemical Research* 37(12) (2012) 2777-2786.
- [44]. S.E. dal Belo, L.R. Gaspar, P.M.B.G. Maia Campos, J.P. Marty, Skin Penetration of Epigallocatechin-3-Gallate and Quercetin from Green Tea and <i>Ginkgo biloba</i> Extracts Vehiculated in Cosmetic Formulations, *Skin Pharmacology and Physiology* 22(6) (2009) 299-304.
- [45]. J.-Y. Kee, Y.-H. Han, D.-S. Kim, J.-G. Mun, J. Park, M.-Y. Jeong, J.-Y. Um, S.-H. Hong, Inhibitory effect of quercetin on colorectal lung metastasis through inducing apoptosis, and suppression of metastatic ability, *Phytomedicine* 23(13) (2016) 1680-1690.
- [46]. G. Sharmila, F.A. Bhat, R. Arunkumar, P. Elumalai, P. Raja Singh, K. Senthilkumar, J. Arunakaran, Chemopreventive effect of quercetin, a natural dietary flavonoid on prostate cancer in in vivo model, *Clinical Nutrition* 33(4) (2014) 718-726.
- [47]. B.L. Santos, A.R. Silva, B.P.S. Pitanga, C.S. Sousa, M.S. Grangeiro, B.O. Fragomeni, P.L.C. Coelho, M.N. Oliveira, N.J. Menezes-Filho, M.F.D. Costa, R.S. El-Bachá, E.S. Vellozo, G.P. Sampaio, S.M. Freire, M. Tardy, S.L. Costa, Antiproliferative, proapoptotic and morphogenic effects of the flavonoid rutin on human glioblastoma cells, *Food Chemistry* 127(2) (2011) 404-411.
- [48]. A.J. Alonso-Castro, F. Domínguez, A. García-Carrancá, Rutin Exerts Antitumor Effects on Nude Mice Bearing SW480 Tumor, *Archives of Medical Research* 44(5) (2013) 346-351.
- [49]. J.R. Araújo, P. Gonçalves, F. Martel, Chemopreventive effect of dietary polyphenols in colorectal cancer cell lines, *Nutrition Research* 31(2) (2011) 77-87.
- [50]. M. Jeszka-Skowron, M. Krawczyk, A. Zgoła-Grześkowiak, Determination of antioxidant activity, rutin, quercetin, phenolic acids and trace elements in tea infusions: Influence of citric acid addition on extraction of metals, *Journal of Food Composition and Analysis* 40 (2015) 70-77.
- [51]. J. Yang, J. Guo, J. Yuan, In vitro antioxidant properties of rutin, *LWT - Food Science and Technology* 41(6) (2008) 1060-1066.
- [52]. S.S. Chen, J. Gong, F.T. Liu, U. Mohammed, Naturally occurring polyphenolic antioxidants modulate IgE-mediated mast cell activation, *Immunology* 100(4) (2000) 471-480.
- [53]. N. Ihme, H. Kiesewetter, F. Jung, K.H. Hoffmann, A. Birk, A. Muller, K.I. Grutzner, Leg oedema protection from a buckwheat herb tea in patients with chronic venous insufficiency: A single centre, randomised, double blind, placebo controlled clinical trial, *Eur. J. Clin. Pharmacol.* 50(6) (1996) 443-447.
- [54]. L. Selloum, H. Bouriche, C. Tigrine, C. Boudoukha, Anti-inflammatory effect of rutin on rat paw oedema, and on neutrophils chemotaxis and degranulation, *Experimental and Toxicologic Pathology* 54(4) (2003) 313-318.
- [55]. H. Javed, M.M. Khan, A. Ahmad, K. Vaibhav, M.E. Ahmad, A. Khan, M. Ashafaq, F. Islam, M.S. Siddiqui, M.M. Safhi, F. Islam, Rutin prevents cognitive impairments by ameliorating oxidative stress and neuroinflammation in rat model of sporadic dementia of Alzheimer type, *Neuroscience* 210 (2012) 340-352.
- [56]. S.-Y. Park, S.-H. Bok, S.-M. Jeon, Y.B. Park, S.-J. Lee, T.-S. Jeong, M.-S. Choi, Effect of rutin and tannic acid supplements on cholesterol metabolism in rats, *Nutrition Research* 22(3) (2002) 283-295.
- [57]. S. Kalganekar, H.B. Gross, W. Yokoyama, C.L. Keen, Effects of a Flavonol-Rich Diet on Select Cardiovascular Parameters in a Golden Syrian Hamster Model, *J. Med. Food* 13(1) (2010) 108-115.
- [58]. T. Koda, Y. Kuroda, H. Imai, Protective effect of rutin against spatial memory impairment induced by trimethyltin in rats, *Nutrition Research* 28(9) (2008) 629-634.
- [59]. M.M. Khan, A. Ahmad, T. Ishrat, G. Khuwaja, P. Srivastawa, M.B. Khan, S.S. Raza, H. Javed, K. Vaibhav, A. Khan, F. Islam, Rutin protects the neural damage induced by transient focal ischemia in rats, *Brain Research* 1292 (2009) 123-135.
- [60]. J. Wu, L. Maoqiang, H. Fan, B. Zhenyu, H. Qifang, W. Xuepeng, Z. Liulong, Rutin attenuates neuroinflammation in spinal cord injury rats, *Journal of Surgical Research* 203(2) (2016) 331-337.
- [61]. S.-w. Wang, Y.-J. Wang, Y.-j. Su, W.-w. Zhou, S.-g. Yang, R. Zhang, M. Zhao, Y.-n. Li, Z.-p. Zhang, D.-w. Zhan, R.-t. Liu, Rutin inhibits  $\beta$ -amyloid aggregation and cytotoxicity, attenuates oxidative stress, and decreases the production of nitric oxide and proinflammatory cytokines, *NeuroToxicology* 33(3) (2012) 482-490.
- [62]. N. Kamalakkannan, P.S.M. Prince, Antihyperglycaemic and antioxidant effect of rutin, a polyphenolic flavonoid, in streptozotocin-induced diabetic Wistar rats, *Basic & Clinical Pharmacology & Toxicology* 98(1) (2006) 97-103.
- [63]. V.D. Kappel, L. Zanatta, B.G. Postal, F.R.M.B. Silva, Rutin potentiates calcium uptake via voltage-dependent calcium channel associated with stimulation of glucose uptake in skeletal muscle, *Archives of Biochemistry and Biophysics* 532(2) (2013) 55-60.
- [64]. I.T. Abdel-Raheem, Gastroprotective Effect of Rutin against Indomethacin-Induced Ulcers in Rats, *Basic & Clinical Pharmacology & Toxicology* 107(3) (2010) 742-750.
- [65]. H.K. Nam, S.H. Hong, K.C. Shin, D.K. Oh, Quercetin production from rutin by a thermostable beta-rutinosidase from *Pyrococcus furiosus*, *Biotechnology letters* 34(3) (2012) 483-9.
- [66]. M. Reinboth, S. Wolfram, G. Abraham, F.R. Ungemach, R. Cermak, Oral bioavailability of quercetin from different quercetin glycosides in dogs, *British Journal of Nutrition* 104(2) (2010) 198-203.
- [67]. C. Manach, A. Scalbert, C. Morand, C. Rémésy, L. Jiménez, Polyphenols: food sources and bioavailability, *The American Journal of Clinical Nutrition* 79(5) (2004) 727-747.
- [68]. S.-C. Shen, W.-R. Lee, H.-Y. Lin, H.-C. Huang, C.-H. Ko, L.-L. Yang, Y.-C. Chen, In vitro and in vivo inhibitory activities of rutin, wogonin, and quercetin on lipopolysaccharide-induced nitric oxide and prostaglandin E2 production, *Eur. J. Pharmacol.* 446(1) (2002) 187-194.

- [69]. D.R. Friend, G.W. Chang, A colon-specific drug-delivery system based on drug glycosides and the glycosidases of colonic bacteria, *Journal of medicinal chemistry* 27(3) (1984) 261-6.
- [70]. P.C.H. Hollman, M. Bijlsman, Y. van Gameren, E.P.J. Cnossen, J.H.M. de Vries, M.B. Katan, The sugar moiety is a major determinant of the absorption of dietary flavonoid glycosides in man, *Free Radical Research* 31(6) (1999) 569-573.
- [71]. I.B. Jaganath, W. Mullen, M.E.J. Lean, C.A. Edwards, A. Crozier, In vitro catabolism of rutin by human fecal bacteria and the antioxidant capacity of its catabolites, *Free Radic. Biol. Med.* 47(8) (2009) 1180-1189.
- [72]. A.M. Aura, K.A. O'Leary, G. Williamson, M. Ojala, M. Bailey, R. Puupponen-Pimiä, A.M. Nuutila, K.M. Oksman-Caldentey, K. Poutanen, Quercetin Derivatives Are Deconjugated and Converted to Hydroxyphenylacetic Acids but Not Methylated by Human Fecal Flora in Vitro, *J. Agric. Food Chem.* 50(6) (2002) 1725-1730.
- [73]. A. Millet, F. Stintzing, I. Merfort, Flavonol quantification and stability of phenolics in fermented extracts from fresh *Betula pendula* leaves, *Journal of Pharmaceutical and Biomedical Analysis* 53(2) (2010) 137-144.
- [74]. S.M. Duckstein, P. Lorenz, F.C. Stintzing, Conversion of Phenolic Constituents in Aqueous *Hamamelis virginiana* Leaf Extracts During Fermentation, *Phytochem. Anal.* 23(6) (2012) 588-597.
- [75]. B. Agnieszka, T. Tomasz, P. Jaroslaw, H. Ewa, Biotransformations of Prenylated Hop Flavonoids for Drug Discovery and Production, *Current Drug Metabolism* 14(10) (2013) 1083-1097.
- [76]. X. Pang, D. Wen, Y. Zhao, C.-Q. Xiong, X.-Q. Wang, L.-Y. Yu, B.-P. Ma, Steroidal saponins obtained by biotransformation of total furostanol glycosides from *Dioscorea zingiberensis* with *Absidia coerulea*, *Carbohydrate Research* 402 (2015) 236-240.
- [77]. A.R. Rechner, M.A. Smith, G. Kuhnle, G.R. Gibson, E.S. Debnam, S.K. Srari, K.P. Moore, C.A. Rice-Evans, Colonic metabolism of dietary polyphenols: influence of structure on microbial fermentation products, *Free radical biology & medicine* 36(2) (2004) 212-25.
- [78]. M.V. Selma, J.C. Espin, F.A. Tomas-Barberan, Interaction between Phenolics and Gut Microbiota: Role in Human Health, *J. Agric. Food Chem.* 57(15) (2009) 6485-6501.
- [79]. D. Delmas, V. Aires, E. Limagne, P. Dutartre, F. Mazué, F. Ghiringhelli, N. Latruffe, Transport, stability, and biological activity of resveratrol, *Annals of the New York Academy of Sciences* 1215(1) (2011) 48-59.
- [80]. J. Ludwig-Müller, L. Jahn, A. Lippert, J. Püschel, A. Walter, Improvement of hairy root cultures and plants by changing biosynthetic pathways leading to pharmaceutical metabolites: Strategies and applications, *Biotechnology Advances* 32(6) (2014) 1168-1179.
- [81]. R. Hosoda, Y. Horio, K. Shimoda, M. Hamada, H. Hamada, H. Hamada, Regioselective Hydroxylation and Glucosylation of Flavanones with Cultured Plant Cells of *Eucalyptus perriniana*, *Nat. Prod. Commun.* 8(7) (2013) 905-906.
- [82]. Q. Yan, W. Zhou, X. Shi, P. Zhou, D. Ju, M. Feng, Biotransformation pathways of ginsenoside Rb1 to compound K by  $\beta$ -glucosidases in fungus *Paecilomyces bainier* sp. 229, *Process Biochemistry* 45(9) (2010) 1550-1556.
- [83]. C.-Y. Chen, Y.-J. Fu, Y.-G. Zu, W. Wang, F.-S. Mu, M. Luo, C.-Y. Li, C.-B. Gu, C.-J. Zhao, Biotransformation of saponins to astragaloside IV from *Radix Astragali* by immobilized *Aspergillus niger*, *Biocatalysis and Agricultural Biotechnology* 2(3) (2013) 196-203.
- [84]. P. Maragkoudakis, T. Nardi, B. Bovo, V. Corich, A. Giacomini, Valorisation of a milk industry by-product as substrate for microbial growth, *Journal of Biotechnology* 150 (2010) S340-S340.
- [85]. P. Fernandes, A. Cruz, B. Angelova, H.M. Pinheiro, J.M.S. Cabral, Microbial conversion of steroid compounds: recent developments, *Enzyme and Microbial Technology* 32(6) (2003) 688-705.
- [86]. L. Ye, C.-Q. Zhou, W. Zhou, P. Zhou, D.-F. Chen, X.-H. Liu, X.-L. Shi, M.-Q. Feng, Biotransformation of ginsenoside Rb1 to ginsenoside Rd by highly substrate-tolerant *Paecilomyces bainier* 229-7, *Bioresource Technology* 101(20) (2010) 7872-7876.
- [87]. A.M. Collins, M.J. Kennedy, Biotransformations and bioconversions in New Zealand: Past endeavours and future potential, *Australas. Biotechnol.* 9(2) (1999) 86-94.
- [88]. M. Miyazawa, H. Ando, Y. Okuno, H. Araki, Biotransformation of isoflavones by *Aspergillus niger*, as biocatalyst, *Journal of Molecular Catalysis B: Enzymatic* 27(2-3) (2004) 91-95.
- [89]. N. Ben Akacha, M. Gargouri, Microbial and enzymatic technologies used for the production of natural aroma compounds: Synthesis, recovery modeling, and bioprocesses, *Food and Bioprocesses* 94 (2015) 675-706.
- [90]. H.F. Rozenbaum, M.L. Patitucci, O.A.C. Antunes, N. Pereira, Jr., Production of aromas and fragrances through microbial oxidation of monoterpenes, *Brazilian Journal of Chemical Engineering* 23(3) (2006) 273-279.
- [91]. M.-E.F. Hegazy, T.A. Mohamed, A.I. ElShamy, A.-E.-H.H. Mohamed, U.A. Mahalel, E.H. Reda, A.M. Shaheen, W.A. Tawfik, A.A. Shahat, K.A. Shams, N.S. Abdel-Azim, F.M. Hammouda, Microbial biotransformation as a tool for drug development based on natural products from mevalonic acid pathway: A review, *Journal of Advanced Research* 6(1) (2015) 17-33.
- [92]. L. Nuraida, A review: Health promoting lactic acid bacteria in traditional Indonesian fermented foods, *Food Science and Human Wellness* 4(2) (2015) 47-55.
- [93]. D. Mamma, E. Kalogeris, D.G. Hatzinikolaou, A. Lekanidou, D. Kekos, B.J. Macris, P. Christakopoulos, Biochemical Characterization of the Multi-enzyme System Produced by *Penicillium decumbens* Grown on Rutin, *Food Biotechnology* 18(1) (2004) 1-18.
- [94]. N.R. Shin, J.S. Moon, S.Y. Shin, L. Li, Y.B. Lee, T.J. Kim, N.S. Han, Isolation and characterization of human intestinal *Enterococcus avium* EFEL009 converting rutin to quercetin, *Letters in applied microbiology* 62(1) (2016) 68-74.
- [95]. V.D. Bokkenheuser, C.H. Shackleton, J. Winter, Hydrolysis of dietary flavonoid glycosides by strains of intestinal *Bacteroides* from humans, *The Biochemical journal* 248(3) (1987) 953-6.
- [96]. J. Beekwilder, D. Marcozzi, S. Vecchi, R. de Vos, P. Janssen, C. Francke, J.V. Vlieg, R.D. Hall, Characterization of Rhamnosidases from *Lactobacillus plantarum* and *Lactobacillus acidophilus*, *Appl. Environ. Microbiol.* 75(11) (2009) 3447-3454.
- [97]. J. Yang, D. Qian, S. Jiang, E.-x. Shang, J. Guo, J.-a. Duan, Identification of rutin deglycosylated metabolites produced by human intestinal bacteria using UPLC-Q-TOF/MS, *Journal of Chromatography B* 898 (2012) 95-100.
- [98]. D.O. Otieno, N.P. Shah, Endogenous  $\beta$ -glucosidase and  $\beta$ -galactosidase activities from selected probiotic micro-organisms and their role in isoflavone biotransformation in soymilk, *J. Appl. Microbiol.* 103(4) (2007) 910-917.
- [99]. J. Beekwilder, D. Marcozzi, S. Vecchi, R. de Vos, P. Janssen, C. Francke, J. van Hylckama Vlieg, R.D. Hall, Characterization of Rhamnosidases from *Lactobacillus plantarum* and *Lactobacillus acidophilus*, *Appl Environ Microbiol* 75(11) (2009) 3447-54.
- [100]. M.F. Iqbal, W.Y. Zhu, Characterization of newly isolated *Lactobacillus delbrueckii*-like strain MF-07 isolated from chicken and its role in isoflavone biotransformation, *FEMS Microbiol. Lett.* 291(2) (2009) 180-187.
- [101]. D. Di Gioia, V. Bregola, I. Aloisio, I. Marotti, G. Dinelli, Biotransformation of common bean and wheat flavonoid glycosides by lactic acid bacteria, *Journal of Biotechnology* 150, Supplement (2010) 340.
- [102]. M.F. Mazzeo, R. Lippolis, A. Sorrentino, S. Liberti, F. Fragnito, R.A. Siciliano, *Lactobacillus acidophilus*-Rutin Interplay Investigated by Proteomics, *PLoS One* 10(11) (2015) e0142376.

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- [103]. D.D. Heeney, M.G. Gareau, M.L. Marco, Intestinal Lactobacillus in health and disease, a driver or just along for the ride?, *Current Opinion in Biotechnology* 49 (2018) 140-147.
- [104]. W. Stahl, H. van den Berg, J. Arthur, A. Bast, J. Dainty, R.M. Faulks, C. Gärtner, G. Haenen, P. Hollman, B. Holst, F.J. Kelly, M. Cristina Polidori, C. Rice-Evans, S. Southon, T. van Vliet, J. Viña-Ribes, G. Williamson, S.B. Astley, Bioavailability and metabolism, *Molecular Aspects of Medicine* 23(1-3) (2002) 39-100.
- [105]. E.M. Hein, K. Rose, G. Van't Slot, A.W. Friedrich, H.U. Humpf, Deconjugation and degradation of flavonol glycosides by pig cecal microbiota characterized by fluorescence in situ hybridization (FISH), *Journal of Agricultural and Food Chemistry* 56(6) (2008) 2281-2290.
- [106]. L. Gayoso, A.-S. Claerbout, M.I. Calvo, R.Y. Cavero, I. Astiasarán, D. Ansorena, Bioaccessibility of rutin, caffeic acid and rosmarinic acid: Influence of the in vitro gastrointestinal digestion models, *Journal of Functional Foods* 26 (2016) 428-438.
- [107]. C. Morand, C. Manach, V. Crespy, C. Remesy, Respective bioavailability of quercetin aglycone and its glycosides in a rat model, *Biofactors* 12(1-4) (2000) 169-174.
- [108]. I.B. Jaganath, W. Mullen, C.A. Edwards, A. Crozier, The relative contribution of the small and large intestine to the absorption and metabolism of rutin in man, *Free Radic. Res.* 40(10) (2006) 1035-1046.
- [109]. H. Yu, Q. Liu, C. Zhang, M. Lu, Y. Fu, W.-T. Im, S.-T. Lee, F. Jin, A new ginsenosidase from *Aspergillus* strain hydrolyzing 20-O-multi-glycoside of PPD ginsenoside, *Process Biochemistry* 44(7) (2009) 772-775.
- [110]. K.M. Tuohy, L. Conterno, M. Gasperotti, R. Viola, Up-regulating the Human Intestinal Microbiome Using Whole Plant Foods, Polyphenols, and/or Fiber, *J. Agric. Food Chem.* 60(36) (2012) 8776-8782.
- [111]. J.-Y. Liu, H.-S. Yu, B. Feng, L.-P. Kang, X. Pang, C.-Q. Xiong, Y. Zhao, C.-M. Li, Y. Zhang, B.-P. Ma, Selective hydrolysis of flavonoid glycosides by *Curvularia lunata*, *Chinese Journal of Natural Medicines* 11(6) (2013) 684-689.
- [112]. J. Boyer, D. Brown, R.H. Liu, Uptake of quercetin and quercetin 3-glucoside from whole onion and apple peel extracts by Caco-2 cell monolayers, *Journal of Agricultural and Food Chemistry* 52(23) (2004) 7172-7179.
- [113]. L.M. Berger, S. Wein, R. Blank, C.C. Metges, S. Wolffram, Bioavailability of the flavonol quercetin in cows after intraruminal application of quercetin aglycone and rutin, *Journal of Dairy Science* 95(9) (2012) 5047-5055.
- [114]. S. Lesser, R. Cermak, S. Wolffram, Bioavailability of quercetin in pigs is influenced by the dietary fat content, *J. Nutr.* 134(6) (2004) 1508-1511.
- [115]. R. Cermak, S. Landgraf, S. Wolffram, The bioavailability of quercetin in pigs depends on the glycoside moiety and on dietary factors, *Journal of Nutrition* 133(9) (2003) 2802-2807.
- [116]. A.J. Day, J.M. Gee, M.S. DuPont, I.T. Johnson, G. Williamson, Absorption of quercetin-3-glucoside and quercetin-4'-glucoside in the rat small intestine: the role of lactase phlorizin hydrolase and the sodium-dependent glucose transporter, *Biochemical Pharmacology* 65(7) (2003) 1199-1206.
- [117]. I.C.W. Arts, A.L.A. Sesink, M. Faassen-Peters, P.C.H. Hollman, The type of sugar moiety is a major determinant of the small intestinal uptake and subsequent biliary excretion of dietary quercetin glycosides, *British Journal of Nutrition* 91(6) (2004) 841-847.